PyBOP^{® 1} AND PyBroP: TWO REAGENTS FOR THE DIFFICULT COUPLING OF THE α,α-DIALKYL AMINO ACID, Aib.

Eric Frérot, Jacques Coste*, Antoine Pantaloni, Marie-Noélle Dufour and Patrick Jouin Centre CNRS-INSERM de Pharmacologie-Endocrinologie, 34094 Montpellier Cédex 5, France

(Received in Belgium 20 September 1990)

Summary The difficult coupling of α -aminoisobutyric acid (Aib) was carried out using PyBOP[®] and PyBroP in a comparative study with BOP and BroP. These reagents gave good results under simple conditions (one pot, rt, 1 h). Coded amino acids could be coupled with Aib using PyBOP under standard conditions of peptide synthesis without racemization whereas the coupling of two Aib residues required PyBroP/DMAP. A fragment containing an Aib C-terminal could be coupled without epimerization of the penultimate residue.

Introduction

Aib (α -aminoisobutyric acid) is a noncoded amino acid present in peptabol antibiotics, which are isolated from fungi² and have a capacity to form voltage-dependent ion channels in bilayer membranes³ The introduction of Aib into a peptide can result in modifications that are of interest conformationally (formation of α or 3₁₀ helices, or β -turns)⁴ and pharmacologically⁵ However, the introduction of Aib into a peptide remains difficult⁶⁻¹¹ The classic reagent DCC (or DCC/HOBt) gives mediocre yields despite long reaction times^{12,13} Other methods have been used, i e mixed anhydrides^{6,7}, active esters^{12,14}, Aib oxazolone⁷, but none of them gives consistently satisfactory results Coupling reagents, such as 2-chloro 4,6-dimethoxy-1,3,5-triazine¹⁵ or diethyl phosphorobromidate¹⁶ have been used in rare cases

An original method using an azirine as the synthetic equivalent of Aib has recently been developed¹¹ Specifically, it amounts to coupling with the Aib amine, which, after two steps, produces a peptide having an acidic Aib C-terminal Further coupling with this carboxyl must be carried out using classical methods¹¹ involving the limitations described above

Because of the difficulty of this coupling and the presence of many Aib residues in peptaibol sequences, these compounds have only rarely been synthesized, and in these cases in solution^{13,17,18} For the same reasons, SPPS (solid-phase peptide synthesis) is unsuitable, although it has been used to obtain alamethicin¹⁰

We recently developed new peptide coupling reagents of the BOP¹⁹ family PyBOP[®] 2, BroP 3, and PyBroP 4. PyBOP²⁰ 2 gives results that are at least as good as BOP, and it avoids the use and formation of carcinogenic HMPA BroP²¹ 3 et PyBroP²² 4 give better results than reagents 1 and 2 when coupling Nmethylated amino acids The results we have obtained when coupling N-methylated amino acids with BroP and PyBroP, as well as the corresponding differences in reactivity observed between reagents 1 and 2, containing the oxybenzotriazole residue, and the brominated reagents 3 and 4, led us to study their behavior during Aib coupling In the present paper, we report results obtained using reagents 1 to 4 in Aib coupling during the synthesis of di- and tripeptides

Results and Discussion

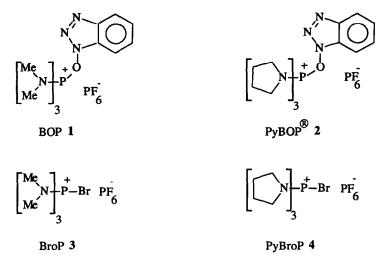
I/ Yields

In a first series of reactions (table I, compounds 5-11), the four reagents 1 to 4 gave high yields (80 to 95% in 1 h of reaction) even with hindered residues (compounds 6-11) The results were identical whether Aib was in the C- or N-terminal position Moreover, there were no differences among the four reagents A second series of reactions (compounds 12-15) involved the difficult coupling of two Aib residues The decrease in reactivity is particularly important with the bulky Boc protective group using

4 REAGENTS(a,b)	
-I HLIM SONTI	
I: Aib COUPL	
TABLE	

BCITTAR	BOP	PyBOP	BroP	PyBroP		E.	Literature
	Yield%	Yield%	Yield%	Yield%	Ref	Yield%	Conditions
5 Z-Aib-Gly-OEt	92	87	89	87	12	55	DCC (for Me ester)
6 Z-Aib-Val-OMe	88	ı	87	84	12	85	AE/Imidazole
7 Z-Val-Aib-OMe	ł	5	96	1			
8 Z.Aib-Pro-OtBu	82	1	84	95(c)	14	5 80	AE/Imidazole DCC/12h (for Me ester)
9 Boc-Pro-Aib-OMe	1	88	ı	83	18	78	MA/1h(-15°C),1h(-5°C)
10 Z-Aib-Ile-OMe	,	80		•			
11 Boc-Ile-Aib-OMe		83	1	1	- -		
12 Z-Aib-Aib-OMe		89(16 h)	ı	77(16h) 80(c.d)	7 14 6	64 89 5	DCC/2 days AE/Imidazole MA/3h(60°C),12h(rr)
13 Boc-Aib-Aib-OMe	80(16h)	86(16h)	25(24 h) 76(d)	25(24 h) 77(d)	8	54	MA/3h(60°C),12h(rt)
15 Boc-Aib-Aib-OBzl	ı	I	65(d)	61(d)	23 13	63 20	BOP-CI DCC/HOBt/2eq./2 days
16 Z-Aib-Benzocaine	30(24h)	,	66 ^(d) (2h)	65 ^(d) (2h)	22 23	0 53	CDI/60h MA/30h
(a)Reactions for 1h at room temperature, except where otherwise stated.	ure, except where o	therwise stated.	(b)See list of abbreviations ¹ . ^(c) Obtained with 20% PyBroP excess. ^(d) With DMAP	viations ¹ . ^(c) Obi	ained with 20	% PyBroP exces	s. ^(d) With DMAP

E. FRÉROT et al.



BroP and PyBroP (compound 13 25%, 24 h) However, with the two latter reagents, catalysis with DMAP

made it possible to obtain good yields in only 1 h of reaction, although synthesis of Boc-Aib-Aib-OBzl 15 remained difficult even with DMAP catalysis For coupling of Z-Aib with a very weakly nucleophilic amine, benzocaine (p-NH₂-Ph-COOEt), the use of BroP(PyBroP)/DMAP made it possible to obtain 66% (65%) amide 16 in 2 h

Comparison of these results with those in the literature shows that reagents 1-4 are clearly more effective than DCC with respect to yield (compound 5), and reaction time (compounds 8,12,15) Compared to active esters in the presence of imidazole, the results are either similar (compound 6) or better (compounds 8,12) The same observation is also true for mixed anhydride methods (compounds 9,12,16 and 13) With regard to other peptide coupling reagents diethyl phosphobromidate¹⁶ gives results (compound 12 85%, 2 h) comparable to those we obtained Dipeptide 15 could not be obtained with BOP-Cl²³, which was also true of CDI in the case of compound 16 ²⁴ Nevertheless, our results were not as good as those obtained by the azirine method. For example, Z-Aib-Aib-OH is obtained in two steps with 91% yield²⁵ whereas PyBroP/DMAP yielded 80% of the corresponding methyl ester 12 Similarly, Z-Val-Aib-OH is obtained with 95% yield¹¹ as opposed to 90% Z-Val-Aib-OMe 7 with BroP. It should be noted, however, that in the azirine method, the precursor has to be prepared beforehand

The reagents we tested showed certain advantages Compounds 2, 3 and 4 are, like BOP, stable solids that are easy to handle Moreover, PyBOP and PyBroP are now commercially available²⁶ The reactions can be carried out one pot, at room temperature C-protected amino acids can be coupled in the form of a free amine (compound 8), chlorhydrate (compounds 5, 6, 7, etc.), tosylate (compound 15), or trifluoroacetate (compound 18) In the case of BroP (and BOP) HMPA is eliminated during the work-up in the acid washes of the reaction mixture With PyBOP and PyBroP, tris(pyrrolidino)phosphine oxide remaining in the crude product is easily eliminated by rapid chromatography on silica gel

II/ Epimerization

It has been demonstrated that when a urethane-protected amino acid is coupled with PyBOP²⁰, there is no epimerization Moreover, when N-methylated amino acids are coupled with BroP and PyBroP, epimerization is undetectable by ¹H-NMR at 360 MHz^{21,22,27}

In the case of Aib, coupling with PyBOP also occurred without epimerization (Table II) The absence of diastereoisomer 24 (eq 3) during the synthesis of tripeptide 20 (eq 1), as measured by HPLC, showed that the coupling of Z-L-Leu with Aib-OAllyl (eq 1, compound 18) did not involve racemization Similarly, the chirality of Z-L-Leu was preserved during coupling with Aib-Pro-OtBu (eq 2, compound 20)

On the other hand, slight epimerization did occur when coupling with BroP and PyBroP, resulting in compounds 18 (eq. 1) and 20 (eq 2) (Table II). Moreover, epimerization was measurable by ¹H-NMR during the coupling of Z-Ile with Aib-benzocaine (eq 4, compound 26) Signals related to α protons of L-Ile and the epimerization product D-allo-Ile are separate²⁸. Thus, epimerization was evaluated at 05% with BroP and was not detectable with PyBroP

We considered it of interest to study the coupling of a peptide fragment containing an Aib at the C-terminal position In peptide synthesis, fragment coupling that extends the peptide in the direction $N \rightarrow C$

Coupling (Compound	PyBOP	BroP	PyBroP	Method
Z-Leu + Aıb-OAllyl	18	0*	15	-	HPLC
Z-Leu + Aıb-Pro-OtBu	2 0	0*	-	1	HPLC
Z-Ile + Aıb-Benzocair	ne 26	-	0.5	0**	NMR

TABLE II % EPIMER BY STEP TO STEP COUPLING

*not detectable on HPLC

** not detectable on ¹H NMR

Equation 1

$$\begin{array}{c} \begin{array}{c} PyBOP\\ Z-L-Leu + Aib-OAllyl\\ 17 \end{array} \xrightarrow{PyBOP} Z-L-Leu-Aib-OAllyl\\ \hline \begin{array}{c} PyBOP\\ Z-L-Leu-Aib-OAllyl\\ \hline \begin{array}{c} Pro-OtBu\\ 19 \end{array} \xrightarrow{PyBOP} Z-L-Leu-Aib-Pro-OtBu\\ 20 \end{array}$$

Equation 3

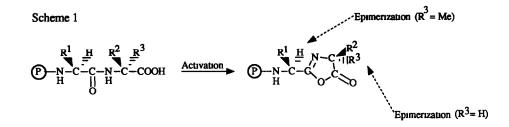
Fountion 2

Z-D-Leu + Aib-OMe $\xrightarrow{\text{BroP}}$ Z-D-Leu-Aib-OMe $\xrightarrow{\text{HO/H}_2\text{O}}$ Z-D-Leu-Aib-OMe $\xrightarrow{\text{Pro-OtBu}}$ Z-D-Leu-Aib-Pro-OtBu 22 23 23 24

Equation 4
Z-Aib-p-NH-C H-COOEt
$$\frac{H_2/Pd/C}{HCl}$$
 HCl, Aib-p-NH-C H-COOEt $\frac{(PyBroP)}{2.64}$ Z-Ile-Aib-p-NH-C H-COOEt $\frac{2.64}{2.64}$ Z-Ile Z-Ile

is used only rarely (except with Gly and Pro C-terminals) because it results in epimerization in the intermediate oxazolone (scheme 1 $R^3 = H$)

If an Aib carboxylate is activated (Scheme 1 $R^2 = R^3 = CH_3$), this epimerization mechanism does not occur and fragment coupling would appear to be advantageous since it allows convergent syntheses However, Heimgartner²⁸ and Bruckner²⁹ have shown that in this case the penultimate amino acid may be epimerized Heimgartner obtained as much as 50% epimer when coupling Z-Ile-Aib with benzocaine using



DCC/HOBt Chirality was preserved when the reaction was carried out in the presence of 10camphorsulfonic acid at 0°C Bruckner proposed a rule he called the "do's and don'ts of Aib peptidecoupling" the coupling of a P-Xxx-Aib fragment is formally forbidden if the chirality of the amino acid Xxx is to be preserved

We studied the coupling of the fragment Z-Leu-Aib with Pro-OtBu (eq 1), a group present in alamethicin This coupling falls into the category of "don'ts" in Bruckner's rule This author obtained as

			-
Reagent	Yield % (2h reaction)	% Epimer (HPLC)	
РуВОР	96	0*	
PyBroP	74	0*	
PyBroP/DMAP	79(1h)	0*	
DCC	86	0*	
DCC/HOBt	85	0*	

TABLE III Z-Leu-Aib + Pro-OtBu COUPLING

*not detectable on HPLC

much as 49 3% epimer while preparing the oxazolone of peptides terminated by Leu-Aib²⁹, which shows the capacity of the Leu fragment to epimerize according to the mechanism considered here

The results in Table III show that epimerization was not detectable under any of the conditions studied This was verified by HPLC in comparison with the diastereoisomer Z-D-Leu-Aib-Pro-OtBu 24 The absence of epimerization with PyBOP and PyBroP (even in the presence of DMAP) led us to carry out the same coupling with DCC and DCC/HOBt Here again, the churality of the Leu residue was preserved These results are in agreement with the low percentages, 1 e 0 7% D-Leu

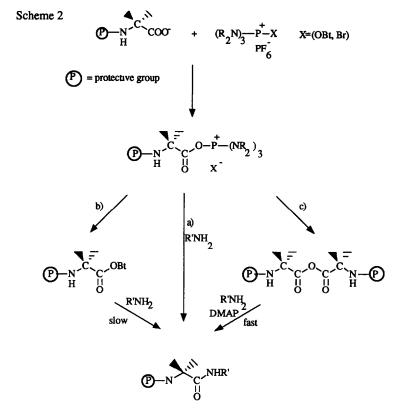
and 0.6% D-Val, present in the alamethicin synthesized by Schmitt and Jung¹⁸ who coupled Aib-terminated fragments with these amino acids in the penultimate position Although it is likely that oxazolone is involved during fragment coupling with Aib C-terminal activation^{17,24}, we do not consider that epimerization of the penultimate residue can occur under the usual conditions of peptide coupling

III/ Mechanism

The very difficult couplings (compounds 12, 13, 15, 16) are of particular interest in terms of the mechanism involved In the cases of Z- and Boc-Aib couplings with Aib-OMe (compounds 12,13) using BOP or PyBOP, thin layer chromatography revealed the immediate formation of an intermediate that slowly disappeared in favor of the dipeptide This kind of intermediate was obtained (TLC) by the action of PyBOP on Boc-Aib Its ¹H-NMR spectrum showed it to be the oxybenzotriazole ester of Boc-Aib 14 During the coupling of Z-Aib with benzocaine (compound 16), the oxybenzotriazole ester of Z-Aib was also formed It reacted even more slowly because of the low nucleophilicity of the amine

In the general case of BOP coupling, we have proposed³⁰ the following mechanism. an acyloxyphosphonium salt is first formed, which is aminolyzed by the C-protected amino acid (Scheme 2,

pathway a) The results given above show that in the case of Aib, if the steric hindrance is large (compounds 12, 13) or the amine is weakly nucleophilic (compound 16), the anion "OBt reacts with the acyloxyphosphonium salt to produce the oxybenzotriazole ester (Scheme 2, pathway b) The latter then reacts slowly to yield the expected product This result is perfectly comparable with those obtained in the coupling of two hindered N-methylated amino acids²¹



For difficult couplings of Aib with BroP and PyBroP, a pathway via a symmetric anhydride can be considered (Scheme 2, pathway c) These reactions are markedly accelerated by DMAP, a reagent known to catalyze the aminolysis of anhydrides (It has also been shown that BroP can be used to prepare anhydrides³¹) This result differs from those obtained^{21,22} with hindered N-methylated amino acids for which DMAP catalysis was unnecessary

Conclusion:

The results obtained and the practical advantages reported here indicate that reagents 1-4 are very suitable for Aib coupling No difference was found between BOP and PyBOP or BroP and PyBroP However, in view of the toxicity of HMPA, PyBOP and PyBroP may be preferable. PyBOP provides good results in most cases and produces peptides that are free of epimerization. In the case of difficult coupling involving two Aib residues, the reaction is slow with PyBOP, but it can be carried out with PyBroP in the presence of DMAP. The results obtained with these two reagents suggest that they can be used in solid-phase synthesis of peptiabols. Studies on this subject are now in progress in our laboratory.

Experimental Section

Melting points are uncorrected ¹H NMR spectra were recorded on a Bruker WM-360 instrument at 360 MHz Optical rotations were taken at 20°C on a Schmidt & Haensch Polartronic D apparatus and are at \pm 1° Elemental analyses were obtained from the "Service Central d'Analyse du CNRS" FAB mass measurements were done by the "Laboratoire de mesures physiques", USTL, Montpelher TLC were performed on silica gel GF254 aluminium sheets (0 2 mm thick; Merck) Column chromatographies were performed using silica gel (0 063-0 200 mm, Merck) Analytical HPLC was carried out on a Beckman System Gold When ¹H NMR and elemental analyses are not given, the products are identical to those previously described in the literature

Coupling methods

BOP, PyBOP, BroP and PyBroP. General procedure: 2 mmol of DIEA (3 mmol if amine salt was present) were added to a solution of 1 mmol N-protected acid component, 1 1 mmol of the C-protected amino acid (or amine salt) and 1 mmol coupling reagent in 1 ml of CH_2Cl_2 (filtered on Alumina) The reaction was stirred at r t for 1 h (except where otherwise stated), evaporated, redissolved in 20 ml of EtOAc and washed 3 x with 5% KHSO₄, brine, 3 x 5% NaHCO₃, brine, dried on Na₂SO₄, filtered and evaporated All dipeptides obtained were purified by column chromatography on silica gel using hexane/EtOAc (except when racemization was studied)

Boc-Aib was synthesized from Aib (3 g) with 3 eq Boc-F (tert-butyl fluorocarbonate, 50% wt in monoglyme, a gift from Propeptide) in 1N NaOH/dioxan (100 ml) at pH=9,5 Dioxan was removed in vacuo, the solution acidified, and the product extracted from EtOAc, washed 3x with brine, dried on Na₂SO₄, evaporated Yield 69% M p =121-122°C (lit ¹² m p = 118°C) Rf = 0.4 (hexane EtOAc, 50.50 1%AcOH)

HCl.H-Aib-OMe was synthesized according to Boissonnas et al ²⁹ From Aib (4 g, 39mmol) Yield 93%. M p = $179-182^{\circ}C$ (ltt ¹² m p = $179-183^{\circ}C$)

Tos.H-Aib-OBzl: A solution of Aib (2 g, 19 mmol), APTS H₂O (3 75 g, 19 mmol) and benzylalcohol (8 ml) in toluene (20 ml) was azeotropically refluxed for 16 h. The colourless solid was filtered and washed with ether Yield 78% M p = $152-153^{\circ}C$ (lit ¹² m p = $154-156^{\circ}C$) Z-Aib-Gly-OEt (5).

With BOP, Z-Aib-OH (0 474 g, 2 mmol), HCl Gly-OEt (0 307 g, 2 2 mmol), (0 884 g, 2 mmol) BOP and **With POP**. From Z Arb-OH (0 474 g, 2 minol), HCl Gly-OE (0 507 g, 2 2 minol), (0 684 g, 2 minol) BOP and DIEA (1 04 ml, 6 mmol) in CH₂Cl₂ (2 ml) Column chromatography (hexane EtOAc, 1 1) Yield 0 57 g (92%). Oil Rf = 0.5 (hexane EtOAc, 1 1) ¹H NMR (CDCl₃) δ 1 25 (3H, t, J = 7 1 Hz, O CH₂CH₃), 1 52 (6H, s, CH₃ Arb), 3 97 (2H, d, J = 4 9 Hz, CH₂ Gly), 4 18 (2H, q, J = 4 1 Hz, OCH₂CH₃), 5 07 (2H, s, CH₂ Z), 5 35 (1H, s, NH Arb), 6 79 (1H, broad s, NH Gly), 7 26-7 35 (5H, m, C₆H₅ Z) Anal calc for C₁₆H₂₂N₂O₅ 1/2 H₂O %C = 57 99, %H = 7 00, %N = 8 45 Found %C = 58 00, %H = 6 85, %N = 8 53 <u>With PyBOP</u>. From Z Arb-OH (2 mmol) Yield 87%

With BroP. From Z-Aib-OH (0 5 mmol) Yield 89% With PyBrOP. From Z-Aib-OH (4 mmol) Yield 87%

Z-Aib-Val-OMe (6):

With BOP: Z-Aib-OH (0 237 g, 1 mmol), HCl H-Val-OMe (0 184 g, 1 1 mmol), BOP (0 442 g, 1 mmol) and DIEA (0 52 ml, 3 mmol) in CH₂Cl₂ (1 ml) Column chromatography (hexane EtOAc, 60 40) Yield 0 294 g (88%) M p = 74°C (ltt ¹² m p = 100-102°C ¹) $[\alpha]_D$ = -3° (c=1, ÉtOH) Rf = 0 55 (hexane EtOAc, 60 40) ¹H NMR (CDCl₃) δ 0 83 (3H, d, J = 6 8 Hz, CH₃ Val), 0 90 (3H, d, J = 6 4 Hz, CH₃ Val), 1 51 (3H, s, CH₃ Aib), 1 55 (3H, s, CH₃ Aib), 2 09-2 18 (1H, m, H- β Val), 3 70 (3H, s, OCH₃), 4 99 (1H, dd, J = 4 9 Hz, J = 8.8 Hz, H- α Val), 5 06-5 10 (2H, AB, J = 12 2 Hz, CH₂ Z), 5 28 (1H, broad s, NH Aib), 6 79 (1H, broad, NH Val), 7 27-7 37 (5H, m, C₆H₅Z) Anal calc for $C_{18}H_{26}N_2O_5$ %C = 61 70, %H = 7 48, %N = 7 99. Found %C =61 90, %H =7 33, %N = 7 79

With BroP, From Z-A1b-OH (0 5 mmol) Yield 87 %

With PyBroP: From Z-Aib-OH (2 mmol) Yield 84%

Z-Val-Aib-OMe (7):

With BroP. Z-Val-OH (0 251 g, 1 mmol), HCl H-Aib-OMe (0 151 g, 1 1 mmol), BroP (0 388 g, 1 mmol) with <u>DiOF</u>, 2-val-OH (0.251 g, 1 minol), the trianely of 0.131 g, 1.1 minol), Brop (0.388 g, 1 minol) and DIEA (0.52 ml, 3 mmol) in CH₂Cl₂ (1 ml) Column chromatography (hexane EtOAc, 60 40) Yield 0.31 g (90%) M p = 83-84°C [α]_D = 24° (c=1, EtOH) ¹H NMR (DMSO d₆) δ 0.83 (3H, d, J = 6.8 Hz, CH₃ Val), 0.86 (3H, d, J = 6.8 Hz, CH₃ Val), 1.32 (3H, s, CH₃ Atb), 1.34 (3H, s, CH₃ Atb), 1.84-1.98 (1H, m, H-β Val), 3.53 (3H, s, OCH₃), 3.85 (1H, dd, J~9 Hz, H-α Val), 5.03 (3H, s, CH₂ Z), 7.10 (1H, d, J = 9.3 Hz, NH Val), 7.27-7.38 (5H, m, C₆H₅ Z), 8.21 (1H, s, NH Atb) Anal calc for C₁₈H₂₆N₂O₅ %C = 61.70, %H = 7.48, %N = 7.99 Found %C = 61.23, %H = 7.56, %N = 7.91

Z-Aib-Pro-OtBu (8):

With BOP Z-A1b-OH (1 896 g, 8 mmol), H-Pro-OtBu (1 505 g, 8 8 mmol), BOP (3 54 g, 8 mmol) and

DIEA (4 6 ml, 27 mmol) in CH₂Cl₂ (8 ml). Column chromatography (hexane: EtOAc, 40 60) Recrystallized from MeOH/H₂O Yield 2 65g (82%) M.p.= 105°C Rf = 0.3 (hexane EtOAc, 50 50) $[\alpha]_D$ = -91° (c=1,EtOH) ¹H NMR (DMSO d₆): δ 1.30 (6H, s, CH₃ Aib), 1.35 (9H, s, tBu), 1.62-1 75 (2H, m, H- β Pro), 1 75-1.96 (2H, m, H- γ Pro), 3.22-3 34 (1H, m, H- δ Pro), 3.60 (1H, broad, H- δ Pro), 4.09 (1H, H- α H- β fro), 1 75-1.96 (2H, m, H- γ Pro), 3.22-3 34 (1H, m, H- δ Pro), 3.60 (2H, m, OX) (2H, broad d, H- α Pro), 4.96, 5 05 (2H, AB, J = 14.4 Hz, CH₂ Z), 7.27-7.38 (5H, m, C₆H₅ Z), 7.70 (1H, s, NH A1b) Anal. calc for C₂₁H₃₀O₅N₂ %C = 64 62, %H = 7.69, %N = 7.18 Found %C = 64.80, %H = 8.14, %N=7.24.

With BroP: From Z-Aib-OH (1 mmol). Yield: 84%.

With PyBroP; From Z-A1b-OH (10 mmol), PyBroP (12 mmol). Yield 95%

Boc-Pro-Aib-OMe (9):

With PyBOP, Boc-Pro-OH (0.43 g, 2 mmol), HCl H-Aib-OMe (0 337 g, 2 2 mmol), PyBOP (1 04 g, 2 mmol) and DIEA (1 04 ml, 6 mmol) in CH₂Cl₂ (2 ml). Column chromatography (hexane:EtOAc; 1.1). Yield 88%. M p = 78°C ($ht.^{18}$ m p = 80°C); Rf = 0.4 (hexane EtOAc, 50.50) $[\alpha]_{D} = -56^{\circ}$ (c=1,EtOH) Anal calc for $\hat{C}_{15}H_{26}N_2O_5$ %C = 57 31, %H = 8 34, %N = 8 91 Found: %C = 57 14; %H = 8 53, %N = 8.89.

With PyBroP. From Boc-Pro-OH (2 mmol) Yield 83% Z-Aib-Ile-OMe (10):

<u>With PyBOP</u>: Z-A1b-OH (0.474 g, 2 mmol), HCl H-Ile-OMe (0 399 g, 2 2 mmol), PyBOP (1 04 g, 2 mmol) and DIEA (1 04 ml, 6 mmol) in CH₂Cl₂ (2 ml) Column chromatography (hexane EtOAc, 50 50). Yield. 80 % M p = 48-50°C. Rf = 0.4 (hexane EtOAc, 1 1) $[\alpha]_D$ = +2° (c=1,EtOH) ¹H NMR (CDCl₃) δ 0.85 (3H, d, J = 6.8 Hz, CH₃ Ile), 0.87 (3H, t, J = 7.3 Hz), 1.04-1.16 (1H, m, H- γ Ile), 1.31-1.43 (1H, m, H- γ Ile), 0.87 (3H, t, J = 7.3 Hz), 0.04 (0.100 Hz) (1 50 (3H, s, CH₃ Aib), 1 53 (3H, s, CH₃ Aib), 1 81-1 92 (1H, m, H-β Ile), 3 68 (3H, s, OCH₃), 4 53 (1H, dd, J = 4.4 Hz, J = 8.3 Hz, $H-\alpha$ Ile), 5.06, 5.08 (2H, AB, J = 12.7 Hz, CH_2 Z), 5.31 (1H, broad s, NH Ab), 6.81 (1H, broad, NH Ile), 7 25-7 48 (5H, m, C₆H₅ Z) Anal calc for $C_{19}H_{28}^2O_5N_2$ %C = 62 64, %H = 7 69, %N = 7 69 Found %C = 62.91, %H = 7 63; %N = 7 72

Boc-Ile-Aib-OMe (11)

With PyBOP, Boc-Ile-OH (0 462 g, 2 mmol), HCl H-Aib-OMe (0 338 g, 2 2 mmol), PyBOP (1 04 g, 2 mmol) and DIEA (1 04 ml, 6 mmol) in CH_2Cl_2 (2 ml) Column chromatography (hexane.EtOAc, 1 1) Yield 83% m p = 168°C. $[\alpha]_{D}$ = -28° (c=1, EtOH) Rf = 0.8 (hexane EtOAc, 50.50). ¹H NMR (CDCl₃) δ 0 88 (3H, t, J = 7 3 Hz, CH₃ Ile), 0 90 (3H, d, J = 6 8 Hz, CH₃ Ile), 1 04-1 17 (1H, m, H-γ Ile), 1.42 (9H, s, CH₃ Boc), 1.40-1 50 (1H, H-γ Ile), 1 507 (3H, s, CH₃ Aib), 1 513 (3H, s, CH₃ Aib), 1.77-1 88 (1H, t, H-β Ile), 3.69 (3H, s, OCH₃), 3 85 (1H, ~t, J ~ 7 3 Hz, H- α Ile), 5 00 (1H, broad, NH Ile), 6.49 (1H, s, NH Ab) Anal calc for $C_{16}H_{30}N_2O_5$ %C = 58 18, %H = 9 09, %N = 8 48 Found %C = 58 54, %H = 9 32, %N = 8 4 4

Z-Aib-Aib-OMe (12) :

With PyBOP, Z-Aib-OH (0 237 g, 1 mmol), HCl H-Aib-OMe (0 169 g, 1 1 mmol), PyBOP (0 52 g, 1 mmol) and DIEA (0 52 ml, 3 mmol) in CH_2Cl_2 (1 ml) TLC monitoring showed immediate formation of one product (Rf = 0 4, hexane EtOAc, 70.30) which disappeared slowly in favor of 12 Reaction time 16 h Column chromatography (hexane EtOAc, 60 40) Yield 89% M p = 106-108°C (lit ⁶ m.p = 107-109°C) Rf = 0.35 (hexane EtOAc, 60.40)

With PyBroP, From Z-Aib-OH (1 mmol) Reaction time 16 h Yield 77%

With PyBroP/DMAP; Z-Aib-OH (0 4587 g, 1 935 mmol), HCl H-Aib-OMe (0 327 g, 2 13 mmol), PyBroP (1.082 g, 2.32 mmol = 1.2 eq), DIEA (0.8 ml, 2.4 eq), DMAP (0.142 g, 1.161 mmol = 0.6 eq) in CH₂Cl₂ (2 ml) Reaction time 1 h Yield 80%

Boc-Aib -Aib-OMe (13):

With BOP: Boc-Aib-OH (0 203 g, 1 mmol), HCl H-Aib-OMe (0 169 g, 1 1 mmol), BOP (0 442 g, 1 mmol) and DIEA (0.51 ml, 3 mmol) in CH_2Cl_2 (1 ml) Immediate formation of 14 (TLC) which disappeared slowly in favor of 13 Reaction time 16 h Column chromatography (hexane EtOAc, 1 1) Yield 80% $M p = 91-92^{\circ}C$ (ltt ¹⁸ m p = 86-87^{\circ}C) Rf = 0.4 (hexane EtOAc, 1.1)

With PvBOP, From Boc-Aib-OH (2 mmol) Immediate formation of 14 Reaction time 16 h Yield 86% With BroP. From Boc-Aib-OH (1 mmol) Reaction time 24 h Yield 25%

With PyBroP, From Boc-Alb-OH (1 mmol) Reaction time. 24 h Yield 25%

With BroP/DMAP. Boc-Aib-OMe (0 203 g, 1 mmol), HCl H-Aib-OMe (0 169 g, 1 1 mmol), BroP (0 388 g,

<u>1 mmol</u>, DIEA (0.408 ml, 2 4 mmol) and DMAP (0.073 g, 0 6 mmol) in CH₂Cl₂ (1 ml) Reaction time 1 h Column chromatography (hexane.EtOAc, 1 1) Yield 76% <u>With PyBroP/DMAP</u>. Boc-Aib-OH (0 322 g, 1 583 mmol), HCl-H-Aib-OMe (0 2673 g, 1.741 mmol), PyBroP (0 885 g, 1 899 mmol), DIEA (0 65 ml, 3 8 mmol = 2 4 eq) and DMAP (0 116 g, 0 94 mmol = 0 6 eq) in CH₂Cl₂ (1 6 ml) Reaction time 1h Column chromatography (hexane EtOAc, 1·1) Yield 77%

Boc-Aib-OBt (14): Boc-Aib-OH (0 203 g, 1 mmol), BOP (0 442 g, 1 mmol) and DIEA (0 34 ml, 2 mmol) in CH₂Cl₂ (1 ml) were stirred at R T TLC (revealed at 254 nm) showed immediate formation of one product. After 30 min, CH₂Cl₂ was evaporated, the residue was dissolved in EtOAc, washed with KHSO₄ 5%, brine, dried on Na₂SO₄, filtered, evaporated. Yield of crude product: 0.24 g (oil). Rf = 0.65 (hexane:EtOAc; 1:1) . ¹H NMR (CDCl₃): δ 1.46 (s) and 1 5(s) (9H, Boc); 1 71 (6H, s, CH₃ Aib); 5 35 (1H, broad s, NH Aib); 7 3-8.1 (4H, OBt).

Boc-Aib-Aib-OBzl (15):

With BroP/DMAP: Boc-Aib-OH (0 812 g, 4 mmol), Tos Aib-OBzl (1 608 g, 4.4 mmol), BroP (1 552 g, 4 mmol), DIEA (1.63 ml, 9 6 mmol = 2.4 eq) and DMAP (0 293 g, 2.4 mmol = 0.6 eq) in CH₂Cl₂ (4 ml) Column chromatography (hexane EtOAc, 1 1) Yield 65% M p = 121° C (lit.¹³: m p. = $121-122^{\circ}$ C) Rf = 0.6 (hexane.EtOAc, 1:1)

<u>With PyBroP/DMAP</u>: From Boc-Aib-OH (1 mmol) Yield 61% Z-Aib-Benzocaīne (16):

With BOP. Z-Aib-OH (0.474 g, 2 mmol), Benzocaine (0 363 g, 2 2 mmol), BOP (0.884 g, 2 mmol) and DIEA (0 68 ml, 4 mmol) in CH₂Cl₂ (2 ml) TLC monitoring showed immediate formation of one product (Rf = 0 4, hexane EtOAc, 70:30) which reacted slowly to give 16 Reaction time. 24 h. Work-up as general procedure Yield of crude product: 1.1 g TLC still showed the same product, benzocaine and 16 ¹H NMR (CDCl₃). about 30% of 16; numerous signals at $\delta = 7$ 2-8 ppm revealed the presence of Z-Aib oxybenzotriazolyl ester.

With BroP/DMAP. Z-Aib-OH (3.92 g, 16 5 mmol), Benzocaine (2 48 g, 15 mmol), BroP (6 40 g, 16.5 mmol), DIEA (2.8 ml, 16.5 mmol) and DMAP (2 02 g, 16 5 mmol) in CH₂Cl₂ (15 ml) Reaction time 2 h Column chromatography (hexane. EtOAc; 60:40). Yield 66% M p.= 126-7°C (lit.²⁴: m p = 131.0-131.2°C). Rf = 0.4 (hexane: EtOAc; 60:40).

With PyBroP/DMAP, From Z-Aib-OH (1 mmol). Reaction time 2h Yield. 65%

Z-L-Leu-Aib-Pro-OtBu (eq.1):

Tfa.H-Aib-OAllyl (17):

a) Boc-Aib-O-Allyl : Boc-Aib-OH (6 7 g, 33 mmmol) were dissolved in DMF (15 ml); allyl bromide (2 86 ml, 33 mmol) and Cesium carbonate (5 376 g, 16.5 mmol) were successively added and the reaction mixture stirred for 2 h at room temperature CsBr was filtered and the solvent removed *in vacuo*. The crude product was dissolved in CH₂Cl₂, washed with water, dried on Na₂SO₄, evaporated and recrystallized from MeOH/H₂O Yield 79% M.p = 50-51°C Rf = 0 7 (hexane EtOAc, 70·30) ¹H NMR (DMSO d_c) δ 1.32 (6H, s, CH₃ Aib), 1 35 (9H, s, CH₃ Boc), 4 51 (2H, d, J = 5 4 Hz, O-CH₂-CH=CH₂), 5 18 (1H, d, J = 10.8 Hz, O-CH₂-CH=CH₂), 5 30 (1H, dd, J = 17.1 Hz, J = 1 9 Hz, O-CH₂-CH=CH₂), 5 18 (1H, m, O-CH₂-CH=CH₂), 7 22 (1H, broad s, NH Aib) Anal calc for C₁₂H₂₁NO₄ %C = 59 24; %H = 8.70, %N = 5.76 Found: %C = 59 22; %H = 8.71; %N = 5.83

b) Tfa-Aib-O-Allyl Boc-Aib-OAllyl (6 g, 24 69 mmol) was treated with trifluoroacetic acid (12 35 ml) for 35 minutes Excess of TFA was removed in vacuo and the product crystallized from ether/hexane Yield 88% M p = 65-68°C. ¹H NMR (DMSO d₆) δ 1 48 (6H, s, CH₃ Aib), 4.71 (2H, dt, J= 59 Hz, J = 1 4 Hz, O-CH₂-CH=CH₂), 5.28 (1H, dt, J = 11 0 Hz, J = 1.2 Hz, O-CH₂-CH=CH₂), 5.36 (1H, dt, J = 19 0 Hz, J = 1.4 Hz, O-CH₂-CH=CH₂), 5.94 (1H, m, O-CH₂-CH=CH₂), 8.51 (3H, NH₃⁺ Aib). Anal. calc for C₉H₁₄NO₄F₃ %C = 42.02; %H = 5.45; %N = 5.45. Found: %C = 42.00, %H = 5.50; %N=5.23 []) Epimerization study with PvBOP.

Z-L-Leu-Aib-O-Allyl (18): 17 (3.115 g, 12.1 mmol), Z-L-Leu-OH (3.57 g, 13.4 mmol), PyBOP (6 41 g, 12.3 mmol), DIEA (6.2 ml, 36.3 mmol) in CH₂Cl₂ (12 ml) Column chromatography (hexane EtOAc; 70:30). Yield: 87%. Rf = 0.3 (hexane EtOAc; 70:30) M p. = 51-52°C. $[\alpha]_D = -25°$. ¹H NMR (DMSO d₆): δ 0.85 (3H, d, J = 6.8 Hz, CH₃ Leu), 0.87 (3H, d, J = 6.8 Hz, CH₃ Leu), 1 34 (3H, s, CH₃ Aib), 1 37 (3H, s, CH₃ Aib), 1 32-1.38 (2H, m, H- β Leu), 1.62 (1H, m, H- γ Leu), 4 06 (1H, ~q, J ~ 8.8 Hz, H- α Leu), 4.46 (2H, d, J = 4.9 Hz, O-CH₂-CH=CH₂), 5 02 (2H, s, CH₂ Z), 5.15 (1H, J = 10.7 Hz, O-CH₂-CH=CH₂), 5.27 (1H, J = 17.6 Hz, O-CH₂-CH=CH₂), 5.83 (1H, m, O-CH₂-CH=CH₂), 7.24 (1H, d, J = 88 Hz, NH Leu), 7.28-7.40 (5H, m, C₆H₅ Z), 8.21 (1H, s, NH Aib). Anal calc. for C₂₁H₃₀N₂O₅: %C = 64 60; %H = 7.74, %N = 7.17, Found %C = 64 69, %H = 7.72, %N = 7.13.

Z-L-Leu-Aib-OH (19): 18 (4.1 g, 10 5 mmol) and morpholin (2 75 ml, 31.5 mmol) in THF (200 ml) were stirred for 5 minutes in the presence of tetrakis-(triphenylphosphine)palladium(0) (1 21 g, 1 05 mmol) according to ³³ THF was evaporated, the yellow product dissolved in EtOAc, washed 3 x with 5% KHSO₄, extracted with 5% NaHCO₃, the aqueous solution was acidified with 6N HCl and extracted with CH₂Cl₂ The product was crystallized from Et₂O/hexane Yield 63% M p =113-115°C [α]_D=-25° (c=1, EtOH) ¹ NMR (DMSO d₆) δ 0.85 (3H, d, J = 64 Hz, CH₃ Leu), 0 87 (3H, d, J = 6.3 Hz, CH₃ Leu), 1 32 (3H, s, CH₃ Aib), 1 34 (3H, s, CH₃ Aib), 1 37-1 46 (2H, m, H- β Leu), 1 54-1.71 (1H, m, H- γ Leu), 4 05 (1H, ~q, J ~ 8 8 Hz, H- α Leu), 5 02 (2H, s, CH₂ Z), 7 26 (1H, d, J = 8 3 Hz, NH Leu), 7.28-7 40 (5H, m, C₆H₅ Z), 7 97 (1H, s, NH Aib), 12.1 (1H, broad s, COOH). Anal calc for C₁₈H₂₆N₂O₅· %C = 61 70, %H = 7 48, %N = 7 99 Found⁶ %C = 61 39; %H = 7 26, %N = 8.08.

Z-L-Leu-Aib-Pro-OtBu (20): 19 (0 175 g, 0 5 mmol), HCl H-Pro-OtBu (0 114 g, 0 55 mmol), PyBOP (0.26 g, 0.5 mmol) and DIEA (0.264 ml, 1.5 mmol) in CH₂Cl₂ (0.5 ml) Reaction time: 2 h. Yield 96% The crude product contained no D-Leu diastereoisomer 24 (Yield and epimerization were studied by HPLC see below)

2) Epimerization study with BroP:

Z-L-Leu-Aib-OAllyl (18): 17 (5.02 g, 20 mmol), Z-L-Leu-OH (5 83 g, 22 mmol), BroP (7 76 g, 20 mmol) and DIEA (10.2 ml, 60 mmol) in CH₂Cl₂ (20 ml) Column chromatography (hexane EtOAc, 70:30) Yield 74% M p = 49-52°C. $[\alpha]_D$ = -25° (c=1, ÉtOH)

Z-L-Leu-Aib-OH (19): 18 (3.0 g, 7 7 mmol), morpholin (2 01 ml, 23 1 mmol) in THF (150 ml) and tetrakis-(triphenylphosphine)palladium(0) (0 92 g, 0 8 mmol) according to 33 Yield 74%. M p.= 115°C $[\alpha]_{D} = -25^{\circ}$ (c=1,EtOH)

Z-L-Leu-Aib-Pro-OtBu (20): From 19 (0 5 mmol) with PyBOP Reaction time 1 h Yield 81 % % diastereoisomer 24: 1 5 % (Yield and epimerization were studied by HPLC see below)

Z-L-Leu-Aib-Pro-OtBu (eq.2):

HCl.H-Aib-Pro-OtBu (21): 8 (25 g, 64 mmol) from PyBroP, Pd/C (05 g) and concentrated HCl (0 534 ml) in MeOH (60 ml) were sturred under H₂ for 1 h 45 After filtration, MeOH was evaporated and the product crystallized from MeOH/ether. Yield. 87%. M p = 160° C (decomposition) [α]_D= -77° (c=1, EtOH) ¹H NMR (DMSO d₆). δ 1 38 (9H, s, CH₃ tBu), 1 56 (3H, s, CH₃ Aib), 1 59 (3H, s, CH₃ Aib), 1 72-1 82 (1H, m, Pro), 1.84-1 98 (2H, m, Pro), 2 07-2 17 (1H, m, Pro), 3 65 (2H, m, H-δ Pro), 4 25 (1H, m, H- α Pro), 8 41 (3H, s, NH₃⁺ Aib) Anal calc for C₁₃H₂₄N₂O₃Cl⁻%C = 53.51, %H = 8 29, %N = 9 60, %Cl = 12 15 Found. %C = 53.71, %H = 8 59, %N = 9 56, %Cl = 11 87

1) Epimerization study with PyBOP:

Z-L-Leu-Aib-Pro-OtBu (20): 21 (0 585 g, 2 mmol), Z-L-Leu-OH (0 584 g, 2 2 mmol), PyBOP (1 144 g, 2 mmol) and DIEA (1 02 ml, 6 mmol) in CH₂Cl₂ (2 ml) The crude product contained no Z-D-Leu-Alb-Pro-OtBu 24 (HPLC: see below) Column chromatography (hexane EtOAc, 1:1) Yield: 81% $M p = 150-152^{\circ}C [\alpha]_{D} = -89^{\circ} (c=0.7, EtOH) Rf = 0.40$ (hexane EtOAc, 50.50) (ht ^{11.} M p = 144-145°C, $[\alpha]_{D} = -82.2$ (c=0.7; EtOH)). ¹H NMR (DMSO d₆) $\delta 0.88$ (6H, 2d, J = 7.4 Hz, CH₃ Leu), 1.29, 1.31, 1.36 (15H, 3s, CH₃ Aib, tBu), 1.39-1.50 (2H, m, H- β Leu), 1.54-1.87 (5H, 2m, H- β Pro, H- γ Pro, H- γ Leu), 3 49 (2H, m, H-δ Pro), 4.07 (2H, m, H-α Pro, H-α Leu), 4 97, 5 06 (2H, AB, J = 12.6 Hz, CH₂Z), 7 26-7 46 (6H, m, C₆H₅Z, NH Leu), 8 24 (1H, s, NH Aib)

2) Epimerization study with PyBroP: Z-L-Leu-Aib-Pro-OtBu (20): 21 (0 585 g, 2 mmol), Z-Leu-OH (0 584 g, 2 2 mmol), PyBroP (1 03 g, 2.2 mmol) and DIEA (1.02 ml, 6 mmol) in CH₂Cl₂ (2 ml) The crude product contained 1% Z-D-Leu-Alb-Pro-OtBu 24 (for HPLC assay see below) Column chromatography (hexane EtOAc, 1 1) Yield 77% M p = 149-151°C $[\alpha]_D = -89°$ (c=07, EtOH)

Z-D-Leu-Aib-Pro-OtBu (24) (eq.3):

Z-D-Leu-Aib-OMe (22): Z-D-Leu-OH (4 1 g, 15 47 mmol), HCl H-Aib-OMe (3 4 g, 22 mmol), BroP (7 76 g, 20 mmol) and DIEA (10 2 ml, 60 mmol) in CH_2Cl_2 (20 ml) Column chromatography (hexane.EtOAc, 70 30) Yield 75% M p = 86-87°C [α]_D = +25° (c=1, EtOH) Anal calc for $C_{19}H_{28}N_2O_5$ %C = 62 64, %H = 774, %N = 7 64 Found %C = 62 31, %H = 7 86, %N = 7 69 Anal calc for

Z-D-Leu-Aib-OH (23): 22 (2 07 g, 5 68 mmol) were sturred with 2N NaOH (5 7 ml) in MeOH (6 5 ml) for 1h The reaction mixture was then solved in EtOAc (100 ml), the carboxylate extracted with H₂O, acidified with 1N HCl and finally extracted with CH₂Cl₂ Yield 72% M p = 110-111°C $[\alpha]_D = +25^\circ$ (c=1, EtOH)

Z-D-Leu-Aib-Pro-OtBu (24): 23 (0 35 g, 1 mmol), H-Pro-OtBu (0 197 g, 1 1 mmol), BroP (0 388 g, 1 mmol) and DIEA (0.34 ml, 2 mmol) in CH₂Cl₂ (1 ml) Reaction time 1 h The crude product contained 1 3% Z-L-Leu-Aib-Pro-OtBu 20 (HPLC "see below) Column chromatography (hexane EtOAc, 60 40) Yield 70% The product still contained 0 3% D-isomer after purification M p = 150-151°C $[\alpha]_D$ = $^{-36^{\circ}}$ (c=1, EtOH) Rf = 0.36 (hexane EtOAc, 50.50) ¹H NMR (DMSO d₆) $^{-5}$ 0.85 (3H, d, J = 7.4 Hz, CH₃ Leu), 0.87 (3H, d, J = 6.6 Hz, CH₃ Leu), 1.32 (3H, s, CH₃ Aib), 1.34 (3H, s, CH₃ Aib), 1.36 (9H, s, tBu), 1.30-1.54 (2H, broad, H-β Leu), 1.54-2.03 (5H, 3 m, H-β Pro, H-γ Pro, H-γ Leu), 3.44 (2H, m, H-δ Pro), 4.06 (2H, m, H-α Pro, H-α Leu), 5.00, 5.06 (2H, AB, J = 12.5 Hz, CH₂Z), 7.27-7.39 (6H, m, C_H-Z, NH Leu), 7 95 (1H, s, NH A1b) Anal calc for $C_{27}H_{42}N_3O_6$ %C = 64 26, %H = 8 39, %N= 8 33 Found %C = 64 58, %H = 8 58, %N = 8 34.

Z-Ile-Aib-Benzocaine (eq.4):

HCI.H-Aib-Benzocaine (25): 16 (1 921 g, 5 mmol), concentrated HCl (0 625 ml, 7 5 mmol) and Pd-C (150 mg) in MeOH (40 ml) were allowed to react with H_2 The product was recrystallized from

MeOH/Et₂O Yield. 90% M p. = 146-150°C (decomposition) ¹H NMR (CDCl₃): δ 1.31 (3H, t, J = 6 9 Hz, OCH₂CH₂), 1.66 (6H, s, CH₃ Aib), 4 29 (2H, q, J = 6.9 Hz, OCH₂CH₃), 7 88, 795 (4H arom, AA'BB', J = 8.1 Hz), 8 49 (3H, broad s, NH₃⁺ Aib), 10 56 (1H, broad s, NH benzocaine) Anal calc for C₁₃H₁₉N₂O₃Cl. %C = 54 45; %H = 6.68, %N = 9.77, %Cl = 12.36 Found %C = 54.14, %H = 6.71, %N = 9.47, %Cl = 12.31.

Z-Ile-Aib-Benzocaïne (26):

With Brop. Z-IIe-OH (0 265 g, 1 mmol), 25 (0 321 g, 1 1 mmol), BroP (0 388 g, 1 mmol) and DIEA (0 51 ml, 3 mmol) in CH₂Cl₂ (1 ml) Reaction time: 1 h The crude product contained ~0 5% Z-D-allo-IIe-Alb-Benzocaine (NMR) Column chromatography (hexane EtOAc, 1 1) Yield 60% M p = 79-80°C $[\alpha]_{D}$ = +46° (c=1 1,EtOH) (lit.¹¹ m.p.= 78 8-79 5°C, $[\alpha]_{D}$ = 46 4° (c=1 1, EtOH)) Rf = 0 5 (hexane.EtOAc, 1 1) ¹H NMR (CDCl₃) showed: H- α IIe (δ 3 81, t, J = 6 5 Hz) and 0 5% H- α D-allo-IIe (δ 3 95) (see ²⁸) With PyBrop. Z-IIe-OH (0.3076 g, 1.159 mmol), 25 (0 3008 g, 1 049 mmol), PyBrop (0 7458 g, 1 6 mmol) and DIEA (0 51 mmo

and DIEA (0 54 ml, 3.15 mmol) in CH₂Cl₂ (1 05 ml) Reaction time 1 h The crude product contained no D-diastereoisomer (NMR). Column chromatography (hexane EtOAc, 60 40) Yield 60% M p = 77-79°C $[\alpha]_{\rm D} = +47^{\circ}$ (c=1 1, EtOH) Rf = 0.4 (hexane : EtOAc, 60 40) ¹H NMR (CDCl₂) showed H- α Ile (δ 3 81, t, J = 6.5 Hz) and no signal corresponding to D-allo-Ile isomer (δ 3.95 ppm) (see ²⁸) The recording conditions made it possible to confirm that D-isomer was <0.2%

Z-L-Leu-Aib-Pro-OtBu (20): Fragment coupling (table 3):

Z-L-Leu-Aib-OH 19 used for this study was synthesized from PyBOP (see above) Yields and % epimer were measured on the reaction mixture without further work-up

1) Epimerization: The peaks corresponding to the diastereoisomers 20 and 24 were well separated (R T = 5 7 and 6 5 min respectively) using the following conditions

- normal phase column Ultrasphere S1 5µ 250 x 4 6 mm (Beckman)

- 2 ml/min flow of hexane EtOAc, 1 1 (isocratic)

visualisation at 254 nm Integration using Beckman system Gold program

2) Yield: The yields of 20 were determined using the commercially available Z-Pro-Leu-OEt as internal reference under the following conditions:

- Ultrabase C8 5 µ 150 x 4 6 mm column (SFCC)

- visualization at 214 nm

- 2 ml/min flow of acetonitrile water TFA, 50 50 1°/00

Z-Pro-Leu-OEt RT = 39 min, Z-Leu-Aib-Pro-OtBu 20 (and 24) RT = 48 min

Coupling with PyBOP, this reaction is described above (eq 1) Reaction time 2 h Yield 96% No epimerization

With PyBroP: 19 (0 175 g, 0 5 mmol), HCl H-Pro-OtBu (0 114 g, 0 55 mmol), PyBrOP (0 233 g, 0 5 mmol)

and DIEA (0 26 ml, 1 5 mmol) in CH₂Cl₂ (0 5 ml) Reaction time 2 h Yield 74% No epimerization With PyBroP/DMAP. 19 (0 175 g, 0 5 mmol), HCl H-Pro-OtBu (0 114 g, 0 55 mmol), PyBroP (0 233 g, 0 5 mmol), DIEA (0 237 ml, 1 4 mmol) and DMAP (0 189 g, 0 14 mmol) in CH₂Cl₂ (0 5 ml) Reaction time 1 h Yield 79% No epimerization

With DCC. 19 (0 175 g, 0 5 mmol), HCl H-Pro-OtBu (0 114 g, 0 55 mmol), DCC (0 104 g, 0 5 mmol), DIEA (0 264 ml, 1 5 mmol) in CH_2Cl_2 (0 5 ml) Reaction time 5 min at 0°C, 2 h at RT Yield 86% No epimerization

<u>With DCC/HOBt.</u> 19 (0 175 g, 0 5 mmol), HCl H-Pro-OtBu (0 114 g, 0 55 mmol), DCC (0 104 g, 0 5 mmol), HOBt (20% H₂O) (0 0822 g, 0 5 mmol), DIEA (0 264 ml, 1 5 mmol) in CH₂Cl₂ (0 5 ml) Reaction time. 5 min at 0°C, 2 h at RT Yield 85% No epimerization

Acknowlegment

We are indebted to ANVAR for financial support We thank Propertide for a generous gift of Boc-F We are grateful to Prof Bertrand Castro for the interest he showed for this work

References and notes

1-Abbreviations and symbols follow the recommendations of the IUPAC-IUB Joint Commission on Biochemical Nomenclature (Eur J Biochem 1984, 138, 9) In addition the following abbreviations are used **BOP:** (1H-1, 2, 3-benzotriazol-1-yloxy)-tris(dimethylamino)-phosphonium hexafluorophosphate, **PyBOP**[®]: (1H-1, 2, 3-benzotriazol-1-yloxy)-tris(pyrrolidino)-phosphonium hexafluorophosphate, **BroP**: bromo-tris(dimethylamino)-phosphonium hexafluorophosphate, PyBroP: bromo-tris(pyrrolidino)phosphonium hexafluorophosphate, DCC: dicyclohexyl-carbodiimide, BOP-Cl: bis(2-oxo-3oxazolidinyl)phosphinic chloride, HOBt: 1, 2, 3-benzotriazole-1-hydroxyde, DIEA: diisopropylethylamine; DMAP: 4-dimethylaminopyridine, CDI: carbonyl diimidazole, MA: mixed anhydride, AE active ester; Aib: α-aminoisobutyric acid

2-Bodo, B, Rebuffat, S, El Haju, M, Davoust, D J Am Chem Soc 1985, 107, 6011 and references cited therein

3-Muller, P, Rudin, DO Nature 1968, 217, 713

4-Valle, G; Crisma, M, Toniolo, C., Beisswenger, R; Rieker, A, Jung, G J Am Chem Soc 1989, 111, 6128 and references cited therein

5-a)Fauchère, J.L in "Adv. in drug research", Testa, B., Ed, Academic Press, London, 1986, 15, 29

b)Cordopatis, P., Gatos, D, Theodoropoulos, D, Mizrahi, J., Regoli, D, Escher, E. in "Peptides 1984", Ragnarsson, U, Ed, Almqvist and Wiksell International, Stockholm, Sweden, 1984, 349

6-Leplawy, M.T., Jones, D S, Kenner, G W., Sheppard, R C. Tetrahedron 1960, 11, 39 7-Jones, D S, Kenner, G W, Preston, J, Sheppard, R C J Chem Soc 1965, 6227

8-Oekonomopulos, R, Jung, G. Lebigs Ann Chem 1979, 1151. 9-Rich, D.H, Singh, J in "The Peptides", Gross, E, Meienhofer, J, Eds, Academic Press, New York, San Francisco, London, 1979, Vol 1, 241

10-Gisin, B F, Davis, D G; Borowska, Z K, Hall, J.E, Kobayashi, S J Am Chem Soc 1981, 103, 6373 11-Wipf, P, Heimgartner, H Helv Chum Acta 1990, 73, 13 and references cited therein

12-Leibfritz, D, Haupt, E, Dubischar, N, Lachmann, H, Oekonomopoulos, R, Jung, G Tetrahedron 1982, 38, 2165.

13-Balasubramanian, T.M.; Kendrick, N.C.E., Taylor, M., Marshall, G.R., Hall, J.E., Vodyanov, I., Reusser, F J Am Chem Soc 1981, 103, 6127

14-Mc Gahren, WJ., Goodman, M Tetrahedron 1967, 23, 2017

15-Kaminski, Z J Synthesis 1987, 917

16-Gorecka, A, Leplawy, M, Zabrocki, J, Zwierzak, A Synthesis 1978, 474

17-Nagaraj, R , Balaram, P Tetrahedron 1981, 37, 1263

18-Schmitt, H, Jung, G Liebigs Ann Chem. 1985, 321

19-Castro, B., Dormoy, J R , Evin, G , Selve, C Tetrahedron Lett 1975 , 1219

20-Coste, J., Le-Nguyen, D., Castro, B. Tetrahedron Lett 1990, 31, 205
21-Coste, J., Dufour, M.N., Pantaloni, A., Castro, B. Tetrahedron Lett 1990, 31, 669.
22-Castro, B., Coste, J., Dufour, M-N., Pantaloni, A. in "Peptides Chemistry, Stucture and Biology", Rivier, J E., Marshall, G.R. Eds, ESCOM, Leiden, 1990, 900
23-Tung, R.D., Rich, D.H. J. Am. Chem. Soc 1985, 107, 4342
24-Wipf, P., Heimgartner, H. Helv Chim. Acta 1988, 71, 140

25-Obrecht, D, Heimgartner, H Helv Chim Acta 1987, 70, 102 26-PyBOP[®] and PyBroP are available from Novabiochem, Laufelfingen, Switzerland

27-Further studies on epimerization induced by PyBroP are in progress First results display no epimerization (HPLC assays)

28-Wipf, P, Heimgartner, H Helv Chim Acta 1986, 69, 1153

29-Bruckner, H, Currle, M in "Second Forum on Peptides", Aubry, A, Marraud, M, Vitoux, B, Eds Colloque INSERM/John Libbey Eurotext Ltd, 1989, Vol 174, 251

30-Coste, J, Dufour, M-N, Le-Nguyen, D, Castro, B in "Peptides Chemistry, Stucture and Biology", Rivier, JE, Marshall, GR Eds, ESCOM, Leiden, 1990, 885

31-Castro, B, Dormoy, J R Tetrahedron Lett 1973, 3243

32-Boissonas, R A, Guttmann, St, Jaquenoud, P-A, Waller, J-P Helv Chum Acta 1955, 38, 1491

33-Kunz, H, Waldmann, H, Unverzagt, C Int J Pept Protein Res 1985, 26, 493